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**(54) Bioadhesive compositions**

(57) A bioadhesive composition formed by polymerising an aqueous reaction mixture comprising from 5% to 50%, by weight of the reaction mixture, of at least one ionic water soluble monomer, from 10% to 50%, by weight of the reaction mixture, of at least one plasticiser (other than water), from 10% to 50%, by weight of the

reaction mixture, of at least one nonionic water soluble monomer, from 3% to 40%, by weight of the reaction mixture, of water, from 0.05% to 10%, by weight of the reaction mixture, of at least one surfactant and from 1 % to 30%, by weight of the reaction mixture of at least one hydrophobic monomer and/or polymer.

**EP 1 026 219 A1**

## Description

**[0001]** The present invention relates to bioadhesive compositions. One possible application of the compositions of the invention is in the field of biomedical skin electrodes. These electrodes incorporate bioadhesive compositions which are electrically conductive.

**[0002]** Biomedical skin electrodes are widely used in a variety of situations, whenever for example it is required to establish an electrical connection between the surface of the body of a patient and external medical equipment for transmission of electrical signals.

**[0003]** Modern medicine uses many medical procedures where electrical signals or currents are received from or delivered to a patient's body. The interface between medical equipment used in these procedures and the skin of the patient is usually some sort of biomedical electrode. Such electrodes typically include a conductor which must be connected electrically to the equipment, and a conductive medium adhered to or otherwise contacting skin of the patient, and they are of varying types with a wide variety of design configurations which will generally depend on their intended use and whether for example they are to be used as transmission electrodes or sensing i.e. monitoring electrodes.

**[0004]** Among the therapeutic procedures using biomedical electrodes are transcutaneous electric nerve stimulation (TENS) devices used for pain management; neuromuscular stimulation (NMS) used for treating conditions such as scoliosis; defibrillation electrodes to dispense electrical energy to a chest cavity of a mammalian patient to defibrillate heart beats of the patient; and dispersive electrodes to receive electrical energy dispensed into an incision made during electrosurgery.

**[0005]** Among diagnostic procedures using biomedical electrodes are monitors of electrical output from body functions, such as electrocardiograms (ECG) for monitoring heart activity and for diagnosing heart abnormalities.

**[0006]** For each diagnostic, therapeutic, or electrosurgical procedure, at least one biomedical electrode having an ionically conductive medium containing an electrolyte is adhered to or is otherwise contacted with mammalian skin at a location of interest and is also electrically connected to electrical diagnostic, therapeutic, or electrosurgical equipment. A critical component of the biomedical electrode is the conductive medium which serves as the interface between the mammalian skin and the diagnostic, therapeutic, or electrosurgical equipment, and which is usually an ionically conductive medium.

**[0007]** Biomedical electrodes are used among other purposes to monitor and diagnose a patient's cardiovascular activity. Diagnostic electrodes are used to monitor the patient immediately and are only applied to the patient for about five to ten minutes. Monitoring electrodes, however, are used on patients in intensive care for up to three days continuously. In contrast, Holter electrodes are used to monitor a patient during strenuous and daily activities.

**[0008]** Although all of the biomedical electrodes just referred to are used to record cardiovascular activity, each electrode requires specific features or characteristics to be successful. Thus, the diagnostic electrode does not have to remain adhered to a patient for extensive periods but it does have to adhere to hairy, oily, dry and wet skin effectively for the five to ten minutes of use. The monitoring electrode has to adhere for a longer period of time although the patient is often immobile during the monitoring period. The Holter electrodes is susceptible to disruption from adhesion due to physical motion, perspiration, water, etc., and therefore requires the best adhesion and at the same time comfort and electrical performance.

**[0009]** In the biomedical electrodes known in the prior art the ionically conductive medium which serves as an interface, between the skin of a mammalian patient and the electrical instrumentation, ranges from conductive gels and creams to conductive pressure sensitive adhesives. However, while the conductive media can be in the form of pressure sensitive conductive adhesives, for monitoring or Holter biomedical electrodes the use of such conductive adhesives is not generally adequate on their own to maintain adhesion to mammalian skin and additional hypoallergenic and hydrophobic pressure sensitive adhesives may be employed around the conductive medium to provide the required mammalian skin adhesion. U.S. Patent No. 5012810 (Strand et al.) and U.S. Patents Nos. 4527087, 4539996, 4554924 and 4848353 (all Engel) are examples of documents that disclose biomedical electrodes which have a hydrophobic pressure sensitive adhesive surrounding the conductive medium.

**[0010]** In general, a desirable skin electrode is one which maintains good electrical contact with the skin and is free of localised current hot spots, i.e. exhibits uniform conductivity. For example, it has been found that a prior art electrode utilising karaya-gum tends to creep in use and flatten out, exposing skin to possible direct contact with the current distribution member or lead wire. A desirable skin electrode should also usually have a low electrical impedance.

**[0011]** It is an object of this invention to provide hydrogel skin adhesives possessing controlled and predictable adhesive properties which may be readily varied to suit different uses and, in the case of medical electrodes or similar devices, different configurations or applications.

**[0012]** In particular, individual aspects of the invention seek, respectively, to provide hydrogel skin adhesives which provide good adhesion to moist and wet skin and such adhesives for use in biomedical skin electrodes. These hydrogels would be useful for adhesion to skin which is subject to flushing by water or aqueous solutions. Conventional bioad-

hesives generally provide poor adhesion to wet skin.

[0013] Further aspects of the present invention seek, respectively, to provide hydrogel skin adhesives which provide good adhesion to grease-coated skin and such adhesives for use in biomedical skin electrodes. Such hydrogel adhesives would ideally provide good adhesion to various skin types, taking account, for example, of the various skin types of people of different ethnic origin which tend to have secreted thereon varying amounts and differing types of grease. Such hydrogels would also ideally provide good adhesion to skin to which an artificial layer of grease has been applied, for example from moisturising skin creams.

[0014] According to a first aspect of the present invention there is provided a bioadhesive composition formed by polymerising an aqueous reaction mixture comprising from 5% to 50%, by weight of the reaction mixture, of at least one ionic water soluble monomer, from 10% to 50%, by weight of the reaction mixture, of at least one plasticiser (other than water), from 10% to 50%, by weight of the reaction mixture, of at least one non ionic water soluble monomer and from 3% to 40%, by weight of the reaction mixture, of water.

[0015] The compositions of the invention exhibit water stability. For the purposes of the present invention "water stability" is defined as the maintenance of adhesion to skin or another substrate from a level of 50% to more than 100% of the value of the "as made" hydrogel adhesive when the water content of the hydrogel has increased by absorption of water (from the environment external to the hydrogel).

[0016] According to a second aspect of the present invention there is provided a bioadhesive composition exhibiting water stability as defined herein, said composition being formed by polymerising an aqueous reaction mixture comprising at least one ionic water soluble monomer, at least one plasticiser (other than water) and at least one non ionic water soluble monomer.

[0017] The compositions of the invention exhibit surprisingly good adhesion to both dry and moist skin and on subsequent exposure to large amounts of water. In particular, the hydrogels in accordance with the invention generally provide adhesion on dry skin at no less than 0.5 N/cm.

[0018] The compositions of the invention seem to provide good two stage adhesion with a good initial "first stage" adhesion on first contact of the hydrogel with the skin which adhesion increases with time in the "second stage".

[0019] Whilst providing sufficient adhesion it is noted that the hydrogel adhesives of the invention allow for pain free removal from the skin.

[0020] In a preferred embodiment of the invention the ionic monomer comprises an acrylate based monomer selected for its ability to polymerise rapidly in water. Most preferably the ionic monomer comprises at least one of 2-acrylamido-2-methylpropane sulphonic acid, an analogue thereof or one of its salts, for example, an alkali metal salt such as a sodium, potassium or lithium salt. A particularly preferred example of the ionic monomer is 2-acrylamido-2-methylpropane sulphonic acid, commonly known as NaAMPS, available commercially at present from Lubrizol as either a 50% aqueous solution (reference code LZ2405) or a 58% aqueous solution (reference code LZ 2405A). The reaction mixture preferably comprises from 10% to 50%, and ideally from 30% to 50%, by weight of the reaction mixture, of the ionic monomer.

[0021] In a preferred embodiment of the invention the plasticiser comprises any of the following either alone or in combination: at least one polyhydric alcohol (such as glycerol), at least one ester derived therefrom and/or at least one polymeric alcohol (such as polyethylene oxide). Glycerol is the preferred plasticiser. An alternative preferred plasticiser is the ester derived from boric acid and glycerol. The plasticiser is generally used to plasticise the hydrogel compositions in accordance with the invention and control adhesive and electrical properties, for electrically conducting hydrogels. When water is lost from the hydrogel both the adhesive and electrical properties may change deleteriously. The reaction mixture preferably comprises from 15% to 45%, by weight of the reaction mixture, of plasticiser (other than water).

[0022] In a preferred embodiment of the invention the aforesaid nonionic water soluble monomer will comprise at least one of dialkylacrylamide or an analogue thereof. N,N-dimethylacrylamide (NNDMA) and/or an analogue thereof is preferred. The reaction mixture preferably comprises from 15% to 30% and ideally from 15% to 25%, by weight of the reaction mixture, of the nonionic water soluble monomer.

[0023] Conventional crosslinking agents are used to provide the necessary mechanical stability and to control the adhesive properties of the hydrogel. Although hydrogels can be made with suitable adhesive and, when required electrical properties a sufficient amount of a suitable crosslinker must be used; if too little crosslinker is used, converting the material into a completed electrode becomes impossible. The amount of crosslinking agent required will be readily apparent to those skilled in the art such as from 0.05% to 0.2%, by weight of the reaction mixture. Typical crosslinkers include tripropylene glycol diacrylate, ethylene glycol dimethacrylate, triacrylate, polyethylene glycol diacrylate (PEG400 or PEG600), methylene bis acrylamide.

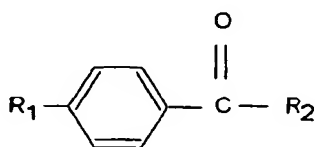
[0024] The adhesives with which this invention is concerned generally comprise, in addition to a crosslinked polymeric network, an aqueous plasticising medium and, optionally, at least one electrolyte, whilst the materials and processing methods used are normally chosen to provide a suitable balance of adhesive and electrical properties for the desired application. In particular, the type of water and its activity together with the rheological properties of the hydrogels will

generally be controlled to produce a balance of pressure sensitive adhesive properties and, when required, electrical properties. One preferred feature of the process used in carrying out the invention is that to achieve the desired adhesive and electrical properties the final amount of water required in the hydrogel is present in the formulation prior to gellation, i.e. no water is removed from the hydrogel after manufacture and less than 10% during manufacture.

[0025] The method of manufacture of the compositions of the invention would generally involve free radical polymerisation and ideally would involve the use of a photoinitiator or a combination of photoinitiation and thermal initiation. Preferably the reaction mixture comprises from 0.02% to 2%, and ideally from 0.02% to 0.2%, by weight of the reaction mixture of photoinitiator. Preferably the reaction mixture comprises from 0.02% to 2%, and ideally from 0.02% to 0.2%, by weight of thermal initiator. Preferred photoinitiators include any of the following either alone or in combination:

[0026] Type I- $\alpha$ -hydroxy-ketones and Benzilidimethyl-ketals e.g. Irgacure 651. These are believed on irradiation to form benzoyl radicals that initiate polymerisation. Photoinitiators of this type that are preferred are those that do not carry substituents in the *para* position of the aromatic ring. Examples include Irgacure 184, and Irgacure 1173 as marketed by Ciba Chemicals

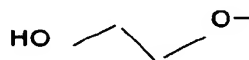
[0027] Photoinitiators of the following general formula are preferred:



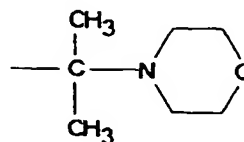
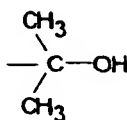
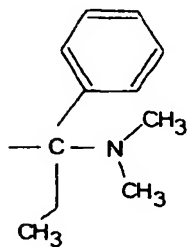
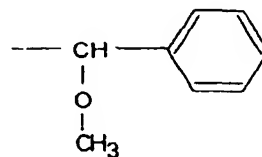
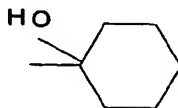
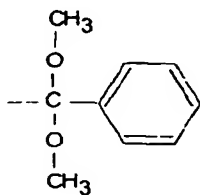
where  $\text{R}_1$  can be any of the following: hydrogen,  $\text{H}_3\text{C-S-}$ ,



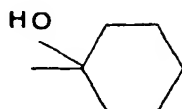
or



$\text{R}_1$  is most preferably hydrogen  
 $\text{R}_2$  can be any of the following:



$\text{R}_2$  is most preferably as follows:-



[0028] A particularly preferred photoinitiator is 1-hydroxycyclohexylphenyl ketone; for example as marketed under the trade name Irgacure 184 by Ciba Speciality Chemicals.

[0029] The adhesives described herein may be used in a range of skin contact applications either unsupported, or in the form of membranes, composites or laminates.

[0030] Such applications include tapes, bandages and dressings of general utility, wound healing and wound management devices; skin contacting, ostomy and related incontinence devices and the like. Other fields of application include pharmaceutical delivery devices, for the delivery of pharmaceuticals or other active agents to or through mammalian skin, optionally containing topical, transdermal or iontophoretic agents and excipients. Non-limiting examples of penetration-enhancing agents include methyl oleic acid, isopropyl myristate, Azone® Transcutol® and N-methyl pyrrolidone.

[0031] A particularly preferred application is in the field of biomedical skin electrodes.

[0032] According to a third aspect of the present invention there is provided the use of a bioadhesive composition in a biomedical skin electrode, said composition being formed by polymerising an aqueous reaction mixture comprising from 5% to 50%, by weight of the reaction mixture, of at least one ionic water soluble monomer, from 10% to 50%, by weight of the reaction mixture, of at least one plasticiser (other than water) from 10% to 50%, by weight of the reaction mixture, of at least one nonionic water soluble monomer, from 3% to 40%, by weight of the reaction mixture, of water and from 0.5% to 6%, by weight of the reaction mixture, of at least one electrolyte.

[0033] According to a fourth aspect of the present invention there is provided the use of a bioadhesive composition in a biomedical skin electrode, said composition exhibiting water stability as defined herein and said composition being formed by polymerising an aqueous reaction mixture comprising at least one ionic water soluble monomer, at least one plasticiser (other than water), at least one non ionic water soluble monomer and at least one electrolyte.

[0034] When the hydrogels are intended for use in conjunction with Ag/AgCl medical electrodes, chloride ions are required to be present in order for the electrode to function. Potassium chloride and sodium chloride are commonly used. However any compound capable of donating chloride ions to the system may be used, for example, lithium chloride, calcium chloride, ammonium chloride. The amount that should be added is dependent on the electrical properties required and is typically 1-7% by weight.

[0035] The main electrical property of interest is the impedance. Performance standards have been drawn up by the American Association of Medical Instruments (AAMI). In sensing electrode applications the electrodes, consisting of the adhesive and a suitable conductive support, are placed in pairs, adhesive to adhesive contact. The conductive support frequently has a Ag/AgCl coating in contact with the adhesive. The measured impedance is dependent on both the quality of the Ag/AgCl coating and the adhesive. In this configuration the adhesive must contain chloride ions. The concentration of chloride ions influences the impedance such that increasing the concentration can lower impedance. It would be anticipated that the activity of the ions (as opposed to the concentration) would be important in determining impedance, but in practice the determination of ion activity in these systems is not a trivial matter. In designing the hydrogel for lowest impedance as measured under the AAMI standard, allowance must be given for the amount and activity of water. These factors will control the effective ion activity and hence the amount of chloride available for participating in the electrochemistry of the system. Hydrogels with lower chloride concentration but higher water activity have lower impedances.

[0036] The present invention further seeks to provide a bioadhesive composition that provides good adhesive performance when applied to naturally greasy skin or artificially grease-coated skin.

[0037] According to a fifth aspect of the present invention there is provided a bioadhesive composition formed by polymerising an aqueous reaction mixture comprising from 5% to 50%, by weight of the reaction mixture, of at least one ionic water soluble monomer, from 10% to 50%, by weight of the reaction mixture, of at least one plasticiser (other than water), from 10% to 50%, by weight of the reaction mixture, of at least one nonionic water soluble monomer, from 3% to 40%, by weight of the reaction mixture, of water, from 0.05% to 10%, by weight of the reaction mixture, of at least one surfactant and from 1 % to 30%, by weight of the reaction mixture of at least one hydrophobic monomer and/or polymer.

[0038] For the avoidance of doubt the term polymer used herein in relation to hydrophobic polymers includes both polymers and copolymers.

[0039] Such compositions exhibit good adhesion to greasy skin. The hydrogels of the invention have been proven by the testing herein to provide adhesion of at least 0.35 N/cm on greasy skin of the type defined in the tests herein.

No residue was left on the skin upon removal.

**[0040]** The invention seeks to provide a homogeneously dispersed reaction mixture comprising both hydrophobic and hydrophilic components which, on polymerisation separates into a biphasic or a multiphasic structure. The phases have in some cases been observed to have a thickness of about 100 microns +/- 50 microns. The reaction mixture contains one or more surface active agents which may assist or promote phase separation but in the course of polymerisation become anisotropically distributed between the resultant phases.

**[0041]** The presence of a hydrophobic monomer or polymer may be necessary in the initial homogenous dispersion in order to more effectively promote phase separation.

**[0042]** It is a consequence of this invention that the phase separated material contains relatively hydrophobic regions, which enable the polymer to function as a pressure sensitive adhesive, and substantially hydrophilic region, which enable the surface active agent to function in an aqueous environment at the interface between the polymer and mammalian skin. When the polymer is placed in contact with skin, the nature and quantity of surface active agent are chosen to bring about the removal of natural or synthetic hydrophobic material, such as skin lipid or skin crème, from the skin surface without adversely diminishing the work of adhesion between the hydrophobic domains and the skin surface. In as much as both the polymeric adhesive formed in this invention and the skin with which it is contacted are deformable under conditions of normal use, an equilibrium interfacial situation is reached in which some spatial exchange of hydrophobic regions and hydrophilic regions will have taken place on the skin surface.

**[0043]** According to a sixth aspect of the present invention there is provided a bioadhesive composition formed by providing a homogeneously dispersed reaction mixture comprising both hydrophobic and hydrophilic components and polymerising said homogeneously dispersed reaction mixture such that on polymerisation the reaction mixture separates into a biphasic or multiphasic structure.

**[0044]** According to a seventh aspect of the present invention there is provided the use of a bioadhesive composition in a biomedical skin electrode, said composition being formed by polymerising an aqueous reaction comprising from 5% to 50%, by weight of the reaction mixture, of at least one ionic water soluble monomer, from 10% to 50%, by weight of the reaction mixture, of at least one plasticiser (other than water), from 10% to 50%, by weight of the reaction mixture, of at least one nonionic water soluble monomer, from 3% to 40%, by weight of the reaction mixture, of water, from 0.05% to 10%, by weight of the reaction mixture, of at least one surfactant, from 1 % to 30%, by weight of the reaction mixture, of at least one hydrophobic monomer and/or polymer and from 0.5% to 6%, by weight of the reaction mixture, of at least one electrolyte.

**[0045]** According to an eighth aspect of the present invention there is provided the use of a bioadhesive composition in a biomedical skin electrode, the biomedical composition being formed by providing a homogeneously dispersed reaction mixture comprising both hydrophobic and hydrophilic components and polymerising said homogeneously dispersed reaction mixture such that on polymerisation the reaction mixture separates into a biphasic or multiphasic structure.

**[0046]** The preferred features of the first, second, third and fourth aspects of the invention as hereinbefore described apply equally to the fifth, sixth, seventh and eighth aspects of the invention.

**[0047]** Any compatible surfactant may be used. Nonionic, anionic and cationic surfactants are preferred. The surfactant ideally comprises any of the surfactants listed below either alone or in combination with other surfactants.

#### 1. Nonionic Surfactants

**[0048]** Suitable nonionic surfactants include but are not limited to those selected from the group consisting of the condensation products of a higher aliphatic alcohol, such as a fatty alcohol, containing about 8 to about 20 carbon atoms, in a straight or branched chain configuration, condensed with about 3 to about 100 moles, preferably about 5 to about 40 moles and most preferably about 5 to about 20 moles of ethylene oxide. Examples of such nonionic ethoxylated fatty alcohol surfactants are the Tergitol.TM. 15-S series from Union. Carbide and Brij.TM. surfactants from ICI. Tergitol.TM. 15-S surfactants include C.sub.11 - C.sub.15 secondary alcohol polyethyleneglycol ethers. Brij.TM. 58 surfactant is polyoxyethylene(20) cetyl ether, and Brij.TM. 76 surfactant is polyoxyethylene(10) stearyl ether.

**[0049]** Other suitable nonionic surfactants include but are not limited to those selected from the group consisting of the polyethylene oxide condensates of one mole of alkyl phenol containing from about 6 to 12 carbon atoms in a straight or branched chain configuration, with about 3 to about 100 moles of ethylene oxide. Examples of nonionic surfactants are the Igepal.TM. CO and CA series from Rhone-Poulenc. Igepal.TM. CO surfactants include nonylphenoxy poly(ethyleneoxy) ethanols. Igepal.TM. CA surfactants include octylphenoxy poly(ethyleneoxy) ethanols.

**[0050]** Another group of usable nonionic surfactants include but are not limited to those selected from the group consisting of block copolymers of ethylene oxide and propylene oxide or butylene oxide.

**[0051]** Examples of such nonionic block copolymer surfactants are the Pluronic.TM. And Tetronic TM. Series of surfactants from BASF. Pluronic.TM. surfactants include ethylene oxide-propylene oxide block copolymers. Tetronic.TM. surfactants include ethylene oxide-propylene oxide block copolymers. The balance of hydrophobic and hydrophilic

components within the surfactant together with the molecular weight are found to be important. Suitable examples are Pluronic L68 and Tetronic 1907. Particularly suitable examples are Pluronic L64 and Tetronic 1107.

[0052] Still other satisfactory nonionic surfactants include but are not limited to those selected from the group consisting of sorbitan fatty acid esters, polyoxyethylene sorbitan fatty acid esters and polyoxyethylene stearates. Examples of such fatty acid ester nonionic surfactants are the Span.TM., Tween.TM., and Myrj.TM. surfactants from ICI. Span.TM. surfactants include C.sub.12 -C.sub.18 sorbitan monoesters. Tween.TM. surfactants include poly(ethylene oxide) C.sub.12 -C.sub.18 sorbitan monoesters. Myrj.TM. surfactants include poly(ethylene oxide) stearates.

## 2. Anionic Surfactants

[0053] Anionic surfactants normally include a hydrophobic moiety selected from the group consisting of (about C. sub.6 to about C.sub.20) alkyl, alkylaryl, and alkenyl groups and an anionic group selected from the group consisting of sulfate, sulfonate, phosphate, polyoxyethylene sulfate, polyoxyethylene sulfonate, polyoxyethylene phosphate and the alkali metal salts, ammonium salts, and tertiary amino salts of such anionic groups.

[0054] Anionic surfactants which can be used in the present invention include but are not limited to those selected from the group consisting of (about C.sub.6 to about C.sub.20) alkyl or alkylaryl sulfates or sulfonates such as sodium lauryl sulfate (commercially available as Polystep.TM B-3 from Stepan Co.) and sodium dodecyl benzene sulfonate, (commercially available as Siponate.TM.DS-10 from Rhone-Poulenc); polyoxyethylene (about C.sub.6 to about C.sub.20) alkyl or alkylphenol ether sulfates with the ethylene oxide repeating unit in the surfactant below about 30 units, preferably below about 20 units, most preferably below about 15 units, such as Polystep.TM.B-1 commercially available from Stepan Co. and Alipal.TM. EP110 and 115 from Rhone-Poulenc; (about C.sub.6 to about C.sub.20) alkyl or alkylphenoxy poly(ethyleneoxy)ethyl mono-esters and di-esters of phosphoric acid and its salts, with the ethylene oxide repeating unit in the surfactant below about 30 units, preferably below about 20 units, most preferably below about 15 units, such as Gafac.TM. RE-510 and Gafac.TM. RE-610 from GAF.

## 3. Cationic Surfactants

[0055] Cationic surfactants useful in the present invention include but are not limited to those selected from the group consisting of quaternary ammonium salts in which at least one higher molecular weight group and two or three lower molecular weight groups are linked to a common nitrogen atom to produce a cation, and wherein the electrically-balancing anion is selected from the group consisting of a halide (bromide, chloride, etc.), acetate, nitrite, and lower alkylsulfate (methosulfate etc.). The higher molecular weight substituent(s) on the nitrogen is/are often (a) higher alkyl group(s), containing about 10 to about 20 carbon atoms, and the lower molecular weight substituents may be lower alkyl of about 1 to about 4 carbon atoms, such as methyl or ethyl, which may be substituted, as with hydroxy, in some instances. One or more of the substituents may include an aryl moiety or may be replaced by an aryl, such as benzyl or phenyl.

[0056] In a particularly preferred embodiment of the invention the surfactant comprises at least one propylene oxide/ethylene oxide block copolymer, for example such as that supplied by BASF Plc under the trade name Pluronic L64. The reaction mixture ideally comprises from 0.1 % to 5%, by weight of the reaction mixture, of surfactant.

[0057] The surfactant acts to remove the grease from the skin and to form the removed grease into isolated pockets within the hydrogel without reducing the work of adhesion of the coating.

[0058] In a preferred embodiment of the invention the homogeneously dispersed reaction mixture preferably further comprises from 0.1 % to 5% by weight of the reaction mixture of a lipid-micellising polymer, i.e. a so-called hypercoiling polymer. This polymer functions to micellise and remove the rolled up pockets of grease from the gel-skin interface.

[0059] This hypercoiling polymer has the capability of more effectively solvating the primary surfactant micelles that contact hydrophobic skin contaminant such as skin lipid or skin cr  me. The consequence of this functional role is that the work of adhesion between adhesive and skin is progressively less affected by the presence of either or both surfactant or hydrophobic skin contaminant.

[0060] The hypercoiling polymer preferably comprises any of the following either alone or in combination: poly (maleic acid-styrene), poly (maleic acid-butyl vinyl ether), poly (maleic acid-propyl vinyl ether), poly (maleic acid-ethyl vinyl ether) and poly (acrylic acid-ethyl acrylate).

[0061] A particularly preferred example is an alternating copolymer of styrene and maleic acid. As stated previously the composition of the invention seeks to provide a biphasic structure on polymerisation. These two phases are hydrophilic and hydrophobic. The hydrophobic phase may be provided by a hydrophobic monomer which is initially maintained as part of the homogenous reaction mixture by way of a reactive solvent bridge as discussed hereinafter. Alternatively and/or additionally the hydrophobic component is provided as a polymer which separates from the aqueous phase on polymerisation.

[0062] The reaction mixture preferably comprises from 1 % to 15%, by weight of the reaction mixture, of a hydrophobic

non-water soluble monomer. This hydrophobic monomer ideally comprises any of the following either alone or in combination: n-butyl acrylate, n-butyl methacrylate, a hexyl acrylate, iso octyl acrylate, isodecyl acrylate, ethoxyethyl acrylate, tetra-hydrofurfuryl acrylate, vinyl propionate and vinyl butyrate. Particularly preferred are ethoxy ethyl acrylate or butyl acrylate. When the hydrogel comprises a hydrophobic component, such as butyl acrylate as well as a hydrophilic monomer (i.e. the aforesaid water soluble ionic monomer), such as NaAMPS, the aforesaid nonionic water soluble monomer, in the preferred example NNDMA, acts as a so-called "reactive solvent bridge" to provide intimate mixing of the various seemingly incompatible components of the reaction mixture prior to polymerisation. The reaction mixture thus has a homogenous structure containing both hydrophilic and hydrophobic components that are intimately mixed, as the NNDMA acts as a solvent for both hydrophilic and hydrophobic materials, providing a clear compatible coating solution or dispersion. As the reactive solvent bridge is polymerised and thus essentially removed from the reaction mixture the stability of the system is adversely affected and the compatible coating solutions or dispersions undergo phase separation so as to provide a biphasic structure.

[0063] In certain circumstances the reaction mixture preferably comprises from 3% to 20%, and more preferably from 8% to 18%, by weight of the reaction mixture, of a stabilised hydrophobic polymer dispersion that is used to provide a stable phase separated system. The hydrophobic polymer preferably comprises any of the following either alone or in combination: vinylacetate dioctyl maleate copolymer or ethylene-vinyl acetate copolymer. Ethylene-vinyl acetate copolymer is preferred, such as that marketed under the trade name DM137 by Harco Chemicals.

[0064] In preparing bioadhesive compositions in accordance with the invention, the ingredients will usually be mixed to provide a reaction mixture in the form of an initial pre-gel aqueous based liquid formulation, and this is then converted into a gel by a free radical polymerisation reaction. This may be achieved for example using conventional thermal initiators and/or photoinitiators or by ionizing radiation. Photoinitiation is a preferred method and will usually be applied by subjecting the pre-gel reaction mixture containing an appropriate photoinitiation agent to UV light after it has been spread or coated as a layer on siliconised release paper or other solid substrate. The incident UV intensity, at a wavelength in the range from 240 to 420nm, is ideally substantially 40mW/cm<sup>2</sup>. The processing will generally be carried out in a controlled manner involving a precise predetermined sequence of mixing and thermal treatment or history.

[0065] The UV irradiation time scale should ideally be less than 60 seconds, and preferably less than 10 seconds to form a gel with better than 95% conversion of the monomers and for conversion better than 99.95% exposure to UV light less than 60 seconds and preferably less than 40 seconds is preferred. Those skilled in the art will appreciate that the extent of irradiation will be dependent on the thickness of the reaction mixture, concentration of photoinitiator and nature of substrate on to which the reaction mixture is coated and the source of UV.

[0066] These timings are for medium pressure mercury arc lamps as the source of UV operating at 100 W/cm. The intensity of UV between 240nm and 420nm reaching the surface of the substrate is at least 200mW/cm<sup>2</sup> as measured on a Solascope from Solatell. For a given lamp UV intensity in a function of the operating power and distance of the reaction mixture from the UV source.

[0067] It is noted that although the adhesives of this invention are normally prepared as sheets, coatings or laminates, other and non limiting forms of preparation include fibres, strands pellets or particles. Particular bio-adhesives, for example may find application in, buccal or gastrointestinal drug delivery systems.

[0068] The invention will be further described with reference to the formulations hereinafter.

[0069] All formulations detailed below were coated onto polyurethane foam (EV1700X from Caligen) at a coat weight of 0.8 to 1.6kg per square meter and cured by exposure to ultraviolet radiation emitted from a medium pressure mercury arc lamp operating at 100 W/cm power for 10 seconds.

#### Example 1

[0070] Mix 6.0g of Irgacure 184 with 20g IRR280(PEG400 diacrylate) from UCB (Solution A). To 0.07g of Irgacure 184 add 23.5g of NNDMA and stir for one hour (keep container covered from light). Add 30g of glycerol to this and stir for 5 minutes, followed by 40g of NaAMPS (58%). Stir for another 5 minutes. Add 0.13g of solution A and stir the whole formulation for 1 hour before use.

#### Example 2

[0071] Mix 6.0 g of Irgacure 184 with 20g IRR280(PEG400 diacrylate) from UCB (Solution A). To 0.07g of Irgacure 184 add 23.5g of NNDMA and stir for one hour (keep container covered from light). Add to this 10g of Mowilith DM137 (50% dispersion of ethylene vinyl acetate copolymer in water from Harco) and stir for 5 minutes. Add 30g of glycerol to this and stir for 5 minutes, followed by 40g of NaAMPS (58%). Stir for another 5 minutes. Add 0.13g of solution A and stir the whole formulation for 1 hour before use.



Example 3

[0072] Mix 6.0g of Irgacure 184 with 20g IRR280(PEG400 diacrylate) from UCB (Solution A). To 0.07g of Irgacure 184 add 23.5g of NNDMA and stir for one hour (keep container covered from light) Add to this 10g of Mowilith DM137 (50% dispersion of ethylene vinyl acetate copolymer in water from Harco) and stir for 5 minutes. Add 30g of glycerol to this and stir for 5 minutes, followed by 40g of NaAMPS (58%). Stir for another 5 minutes. Add 0.5g of Pluronic L64 (poly(ethylene glycol) - block - poly(propylene glycol) - block - poly(ethylene glycol) available from BASF). Add 0.13g of solution A and stir the whole formulation for 1 hour before use.

Example 4

[0073] Mix 6.0g of Irgacure 184 with 20g IRR280(PEG400 diacrylate) from UCB (Solution A). To 0.07g of Irgacure 184 add 23.4g of NNDMA and stir for one hour (keep container covered from light). Add to this 2g of Mowilith DM137 (50% dispersion of ethylene vinyl acetate copolymer in water from Harco) and stir for 5 minutes. Add 36g of glycerol to this and stir for 5 minutes, followed by 40.36g of NaAMPS (58%). Stir for another five minutes. Add 0.25g of Pluronic L64 (poly(ethylene glycol) - block - poly(propylene glycol) - block - poly(ethylene glycol) available from BASF). To this add 0.8g of a 30% aqueous solution of poly(styrene-altmaleic acid) sodium salt available from Aldrich and stir for 10 minutes. Add 0.13g of solution A and stir the whole formulation for 1 hour before use.

Example 5

[0074] Mix 6.0g of Irgacure 184 with 20g IRR280(PEG400 diacrylate) from UCB (Solution A). To 0.07g of Irgacure 184 add 23.4g of NNDMA and stir for one hour (keep container covered from light). Add to this 10g of Mowilith DM137 (50% dispersion of ethylene vinyl acetate copolymer in water from Harco) and stir for 5 minutes. Add 36g of glycerol to this and stir for 5 minutes, followed by 40.36g of NaAMPS (58%). Stir for another 5 minutes. Add 0.25g of Pluronic L64 (poly(ethylene glycol) - block - poly(propylene glycol) - block - poly(ethylene glycol) available from BASF). To this add 0.8g of a 30% aqueous solution of poly(styrene-altmaleic acid) sodium salt available from Aldrich and stir for 10 minutes. Add 0.13g of solution A and stir the whole formulation for 1 hour before use.

Example 6

[0075] Mix 6.0g of Irgacure 184 with 20g IRR280(PEG400 diacrylate) from UCB (Solution A). To 0.07g of Irgacure 184 add 23.4g of NNDMA and stir for one hour (keep container covered from light). Add to this 10.g of Mowilith DM137 (50% dispersion of ethylene vinyl acetate copolymer in water from Harco) and stir for 5 minutes. Add 36g of glycerol to this and stir for 5 minutes, followed by 40.36g of NaAMPS (58%). Stir for another 5 minutes. Add 0.5g of Pluronic L64 (poly(ethylene glycol) - block - poly(propylene glycol) - block - poly(ethylene glycol) available from BASF). To this add 0.8g of a 30% aqueous solution of poly(styrene-alt-maleic acid) sodium salt available from Aldrich and stir for 10 minutes. Add 0.13g of solution A and stir the whole formulation for 1 hour before use.

Example 7

[0076] Mix 6.0g of Irgacure 184 with 20g IRR280(PEG400 diacrylate) from UCB (Solution A). To 0.07g of Irgacure 184 add 23.4g of NNDMA and stir for one hour (keep container covered from light). Add to this 20g of Mowilith DM137 (50% dispersion of ethylene vinyl acetate copolymer in water from Harco) and stir for 5 minutes. Add 36g of glycerol to this and stir for 5 minutes, followed by 40.36g of NaAMPS (58%). Stir for another 5 minutes. Add 0.5g of Pluronic L64 (poly(ethylene glycol) - block - poly(propylene glycol) - block - poly(ethylene glycol) available from BASF). To this add 0.8g of a 30% aqueous solution of poly(styrene-altmaleic acid) sodium salt available from Aldrich and stir for 10 minutes. Add 0.13g of solution A and stir the whole formulation for 1 hour before use.

Example 8

[0077] To 36 parts glycerol, were added 0.5 parts of a 30% aqueous solution of poly(styrene-alt-maleic acid) sodium salt available from Aldrich and 40.4 parts of a 58% solution of the sodium salt of 2-acrylamido-2-methylpropane sul-phonic acid (NaAMPS) (LZ2405A) together with 0.5 parts Pluronic LF64 (BASF), and the solution stirred to ensure uniform mixing. To the solution was added 0.13 parts of solution containing 20 parts of polyethylene glycol diacrylate (PEG600) (product of UCB Chemicals marketed under the trade name designation of Ebacryl 11) in which 6 parts of 1-hydroxycyclohexyl phenyl ketone (product of Ciba and marketed under the trade name designation of Irgacure 184) had been dissolved. A premixed solution of 8 parts butyl acrylate and 15.7 parts N,N-dimethylacrylamide (Kohjin) was

added to that reaction mixture and this final solution cured by exposure to UV light as in example 1. Optical phase contrast microscopy showed that resultant gel to have a regularly phase-segregated surface and enhanced adhesion to skin that had previously treated with skin cream (Nivea).

Table 1

Effect of water uptake on peel adhesion on dry skin for the formulation in Example 1.	
Subject 1	
% Water uptake	Peel Adhesion (N/cm)
0	1.8
9	2.2
10	2.3
24	1.6

Subject 2	
% Water uptake	Peel Adhesion (N/cm)
0	1.6
9	2.9
11	2.5
12	2.6

Table 2

Effect on peel adhesion on dry skin from the addition of phase separator to the formulation in Example 1		
Peel Strength (N/cm)		
	Example 1	Example 2
Subject 1	1.8	2.9
Subject 2	1.6	3.2

Table 3

Effect of the addition surfactant to the formulation in example 2 on dry and greasy skin.			
Peel Strength (N/cm)			
Greasy Skin			
	Dry	1 min	10 min
Subject 1	2.8	0.52	0.33
Subject 2	2.5	0.67	0.61

Table 4

Peel strength on dry and greasy skin for examples 4,5,6 & 7 Peel Strength (N/cm)						
Subject 1			Subject 2			
Example	Dry	Greasy		Dry	Greasy	
		1 min	10 min		1 min	10 min
4	0.81	0.15	0.26	0.96	0.29	0.47
5	1.2	0.52	0.69	2.2	0.83	0.88
6	1.6	0.45	0.6	2.2	0.64	0.56
7	1.2	0.49	0.62	1.6	0.74	0.88

### Peel Adhesion Method

[0078] This is a method to determine the peel strength required of adhered hydrogel to the skin of two male subjects of different ethnic origin. The skin is tested "dry" (i.e. normal to the subject) and greasy as described next.

Equipment	
Scissors	Convenient source
Standard ruler	Convenient source
Compression weight	5.0 kg, diameter 130mm
Polyester Film	PET 23 $\mu$ available from EFFEGIDI S.p.A.43052 Colomo, Italy
Transfer Adhesive	3M 1524 available from 3M Italia S.p.A. 20090 Segrate, Italy
Stop Watch	Convenient source
Tensile Tester	Instron mod: 6021 (or equivalent)

### Test procedure

#### A) Tensile Tester Peel Settings:-

[0079]

Load cell	10N
Test Speed	1000 mm/min
Clamp to Clamp Distance	25mm
Pre Loading	0.2N
Test Path "LM"	50mm
Measure variable	F average (N) in "LM"

#### B) Sample preparation

[0080]

1. Allow the samples to adjust to conditioned room ( $23 \pm 2^\circ$  Celsius and  $50 \pm 2\%RH$ ) for about 1 hour.
2. Each test specimen should be prepared individually and tested immediately.
3. Prepare rectangular adhesive samples  $100\text{mm} \pm 2$  length and 25.4mm width.
4. On the forearm draw a rectangle about 2cm wider and longer of the area of the glue extrusion. Ring one cotton disk (i.e. Demak up diameters 5.5cm, weight about 0.6g) put on its 4 drops (about 0.20g) of cream "Nivea body" (for normal skin) folding the cotton disk twice in order so that the cotton absorbs the cream and with a light pressure

rub the forearm surface with the treated cotton side three times.

5. Attach adhesive specimen to the forearm within marked area with light pressure.

6. Gently roll the compression weight down the forearm, on the adhesion sample.

7. Remove the weight and test after 1 and 10 minutes by attaching one end of the specimen into the upper jaws of an adhesion testing machine at an initial angle of 90°.

[0081] The same procedure as above is carried out in order to determine the peel strength of the adhesive after absorption of water. The specimen is placed into an oven at 37°C and at 85% humidity. The time of exposure is dependant on the degree of water uptake required. The sample is then removed from the oven and the steps 5 to 7 are carried out.

## Claims

1. A bioadhesive composition formed by polymerising an aqueous reaction mixture comprising from 5% to 50%, by weight of the reaction mixture, of at least one ionic water soluble monomer, from 10% to 50%, by weight of the reaction mixture, of at least one plasticiser (other than water), from 10% to 50%, by weight of the reaction mixture, of at least one non ionic water soluble monomer and from 3% to 40%, by weight of the reaction mixture, of water.
2. A bioadhesive composition exhibiting water stability as defined herein, said composition being formed by polymerising an aqueous reaction mixture comprising at least one ionic water soluble monomer, at least one plasticiser (other than water) and at least one non ionic water soluble monomer.
3. A bioadhesive composition as claimed in claim 2, characterised in that the aqueous reaction mixture comprises from 5% to 50%, by weight of the reaction mixture, of said ionic water soluble monomer.
4. A bioadhesive composition as claimed in claim 2 or claim 3, characterised in that the aqueous reaction mixture comprises from 10% to 50%, by weight of the reaction mixture, of said plasticiser (other than water).
5. A bioadhesive composition as claimed in any of claims 2 to 4, characterised in that the aqueous reaction mixture comprises from 10% to 50%, by weight of the reaction mixture, of said non ionic water soluble monomer.
6. A bioadhesive composition as claimed in any of claims 2 to 5, characterised in that the aqueous reaction mixture comprises from 3% to 40%, by weight of the reaction mixture, of water.
7. A bioadhesive composition as claimed in any preceding claim, characterised in that the composition provides adhesion on dry skin at no less than 0.5 N/cm.
8. A bioadhesive composition as claimed in any preceding claim, characterised in that said ionic monomer comprises an acrylate based monomer.
9. A bioadhesive composition as claimed in any preceding claim, characterised in that said ionic monomer comprises any of 2-acrylamido-2-methylpropane sulphonic acid, an analogue thereof or a salt thereof.
10. A bioadhesive composition as claimed in any preceding claim, characterised in that the reaction mixture comprises from 30% to 50%, by weight of the reaction mixture, of said ionic monomer.
11. A bioadhesive composition as claimed in any preceding claim, characterised in that said plasticiser comprises any of the following either alone or in combination: at least one polyhydric alcohol, at least one ester derived from polyhydric alcohol and at least one polymeric alcohol.
12. A bioadhesive composition as claimed in any preceding claim, characterised in that said plasticiser comprises at least one of glycerol and an ester derived from boric acid and glycerol.
13. A bioadhesive composition as claimed in any preceding claim, characterised in that the reaction mixture comprises from 15% to 45%, by weight of the reaction mixture, of said plasticiser (other than water).
14. A bioadhesive composition as claimed in any preceding claim, characterised in that said nonionic water soluble

monomer comprises at least one of a dialkylacrylamide or an analogue thereof.

15. A bioadhesive composition as claimed in any preceding claim, characterised in that said nonionic water soluble monomer comprises at least one of N,N-dimethylacrylamide or an analogue thereof.

16. A bioadhesive composition as claimed in any preceding claim, characterised in that the reaction mixture comprises from 15% to 25%, by weight of the reaction mixture, of said nonionic water soluble monomer.

17. A bioadhesive composition as claimed in any preceding claim, characterised in that the composition further comprises at least one electrolyte.

18. The use of a bioadhesive composition as claimed in any of claims 1 to 17 in a biomedical skin electrode.

19. A bioadhesive composition formed by polymerising an aqueous reaction mixture comprising from 5% to 50%, by weight of the reaction mixture, of at least one ionic water soluble monomer, from 10% to 50%, by weight of the reaction mixture, of at least one plasticiser (other than water), from 10% to 50%, by weight of the reaction mixture, of at least one nonionic water soluble monomer, from 3% to 40%, by weight of the reaction mixture, of water, from 0.05% to 10%, by weight of the reaction mixture, of at least one surfactant and from 1 % to 30%, by weight of the reaction mixture of at least one hydrophobic monomer and/or polymer.

20. A bioadhesive composition formed by providing a homogeneously dispersed reaction mixture comprising both hydrophobic and hydrophilic components and polymerising said homogeneously dispersed reaction mixture such that on polymerisation the reaction mixture separates into a biphasic or multiphasic structure.

21. A bioadhesive composition as claimed in claim 21, wherein the bioadhesive composition comprises effective amounts of at least one ionic water soluble monomer, at least one plasticiser (other than water), at least one nonionic water soluble monomer, water, at least one surfactant, and at least one hydrophobic monomer and/or polymer.

22. A bioadhesive composition as claimed in claim 20 or claim 21, wherein the said reaction mixture comprises from 5% to 50% by weight of the reaction mixture of at least one ionic water soluble monomer, from 10% to 50%, by weight of the reaction mixture, of at least one plasticiser (other than water), from 10% to 50%, by weight of the reaction mixture, of at least one nonionic water soluble monomer, from 3% to 40%, by weight of the reaction mixture, of water, from 0.05% to 10%, by weight of the reaction mixture, of at least one surfactant and from 1 % to 30%, by weight of the reaction mixture of at least one hydrophobic monomer and/or polymer.

23. A bioadhesive composition as claimed in any of claims 19, 21 or 22, characterised in that the composition provides adhesion of at least 0.35 N/cm on greasy skin of the type defined in tests herein.

24. A bioadhesive composition as claimed in any of claims 19 or 21 to 23, characterised in that the composition provides adhesion on dry skin at no less than 0.5 N/cm.

25. A bioadhesive composition as claimed in any of claims 19 or 21 to 24, characterised in that said ionic monomer comprises an acrylate based monomer.

26. A bioadhesive composition as claimed in any of claims 19 or 21 to 25, characterised in that said ionic monomer comprises any of 2-acrylamido-2-methylpropanesulphonic acid, an analogue thereof or a salt thereof.

27. A bioadhesive composition as claimed in any of claims 19 or 21 to 26, characterised in that the reaction mixture comprises from 30% to 50%, by weight of the reaction mixture, of said ionic monomer.

28. A bioadhesive composition as claimed in any of claims 19 or 21 to 27, characterised in that said plasticiser comprises any of the following either alone or in combination: at least one polyhydric alcohol, at least one ester derived from polyhydric alcohol and at least one polymeric alcohol.

29. A bioadhesive composition as claimed in any of claims 19 or 21 to 28, characterised in that said plasticiser comprises at least one of glycerol and an ester derived from boric acid and glycerol.

30. A bioadhesive composition as claimed in any of claims 19 or 21 to 29, characterised in that the bioadhesive composition comprises from 15% to 45%, by weight of the reaction mixture of said plasticiser (other than water).
- 5 31. A bioadhesive composition as claimed in any of claims 19 or 21 to 30, characterised in that the nonionic water soluble monomer comprises at least one of a dialkylacrylamide or an analogue thereof.
32. A bioadhesive composition as claimed in any of claims 19 or 21 to 31, characterised in that said nonionic water soluble monomer comprises at least one of N,N-dimethylacrylamide or an analogue thereof.
- 10 33. A bioadhesive composition as claimed in any of claims 19 or 21 to 32, characterised in that the reaction mixture comprises from 15% to 25%, by weight of the reaction mixture, of said nonionic water soluble monomer.
34. A bioadhesive composition as claimed in any of claims 19 or 21 to 33, characterised in that the reaction mixture comprises from 0.1 % to 5%, by weight of the reaction mixture, of said surfactant.
- 15 35. A bioadhesive composition as claimed in any of claims 19 or 21 to 34, characterised in that said surfactant comprises one or more nonionic surfactants.
36. A bioadhesive composition as claimed in any of claims 19 or 21 to 35, characterised in that the surfactant comprises one or more anionic surfactants.
- 20 37. A bioadhesive composition as claimed in any of claims 19 or 21 to 36, characterised in that the surfactant comprises one or more cationic surfactants.
- 25 38. A bioadhesive composition as claimed in any of claims 19 or 21 to 37, characterised in that the surfactant comprises at least one propylene oxide/ethylene oxide block copolymer.
39. A bioadhesive composition as claimed in any of claims 19 or 21 to 38, characterised in that the reaction mixture further comprises at least one lipid micellising polymer.
- 30 40. A bioadhesive composition as claimed in claim 39, characterised in that the reaction mixture comprises from 0.1 % to 5%, by weight of the reaction mixture, of lipid micellising polymer.
- 35 41. A bioadhesive composition as claimed in claim 39 or claim 40, characterised in that the lipid micellising polymer comprises any of the following either alone or in combination: poly (maleic acid-styrene), poly (maleic acid-butyl vinyl ether), poly (maleic acid-propyl vinyl ether), poly (maleic acid-ethyl vinyl ether) and poly (acrylic acid-ethyl acrylate).
- 40 42. A bioadhesive composition as claimed in any of claims 39 to 41, characterised in that the lipid micellising polymer comprises an alternating copolymer of styrene and maleic acid.
43. A bioadhesive composition as claimed in any of claims 19 or 21 to 42, characterised in that the reaction mixture comprises from 1 % to 15%, by weight of the reaction mixture, of said hydrophobic monomer.
- 45 44. A bioadhesive composition as claimed in any of claims 19 or 21 to 43, characterised in that said hydrophobic monomer comprises any of the following either alone or in combination: n-butyl acrylate, n-butyl methacrylate, a hexyl acrylate, iso octyl acrylate, isodecyl acrylate, ethoxyethyl acrylate tetrahydrofurfuryl acrylate, vinyl propionate, and vinyl butyrate.
- 50 45. A bioadhesive composition as claimed in any of claims 19 or 21 to 44, characterised in that the hydrophobic monomer comprises at least one of ethoxy ethyl acrylate or butyl acrylate.
46. A bioadhesive composition as claimed in any of claims 19 or 21 to 45, characterised in that the reaction mixture from 3% to 20%, by weight of the reaction mixture, of said hydrophobic polymer.
- 55 47. A bioadhesive composition as claimed in any of claims 19 or 21 to 46, characterised in that the said hydrophobic polymer comprises any of the following either alone or in combination: vinylacetate dioctyl maleate copolymer or ethylene vinylacetate copolymer.

48. The use of a bioadhesive composition as claimed in any of claims 19 to 47 in a biomedical skin electrode.

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# EUROPEAN SEARCH REPORT

Application Number  
EP 99 30 0740

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION
X	WO 95 20634 A (MINNESOTA MINING & MFG) 3 August 1995 (1995-08-03)  * claims 1-3,9 * * page 14, line 20 - line 28 * * page 15, line 18 * * page 22, line 5 - line 14 * ---	1-10, 13-27, 30-38, 43-48	C09J201/02 A61B5/0408
X	WO 97 05171 A (MINNESOTA MINING & MFG) 13 February 1997 (1997-02-13)  * claims 1,2,9,13,15,20 * * page 15, line 9 - line 17 * * page 16, line 7 * * page 23, line 2 - line 11 * ---	1-10, 13-27, 30-38, 43-48	
X	EP 0 188 381 A (KYOWA GAS CHEM IND CO LTD) 23 July 1986 (1986-07-23)  * claims * * page 4, column 4, line 16 - line 51 * * example 3 * ---	1-6, 8-13, 16-18	TECHNICAL FIELDS SEARCHED
X	WO 81 02097 A (MINNESOTA MINING & MFG) 6 August 1981 (1981-08-06) * claims 1,3-5 * ---	1-6,8, 10-13	C09J A61B A61L
A	EP 0 012 402 A (MEDTRONIC INC) 25 June 1980 (1980-06-25) * claims 1,2 * -----	1	
The present search report has been drawn up for all claims			
Place of search <b>THE HAGUE</b>		Date of completion of the search <b>11 August 1999</b>	Examiner <b>Niaounakis, M</b>
<p><b>CATEGORY OF CITED DOCUMENTS</b></p> <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p> <p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons &amp; : member of the same patent family, corresponding document</p>			

EPO-KOM 1503 03 02 (RevC01)



ANNEX TO THE EUROPEAN SEARCH REPORT  
ON EUROPEAN PATENT APPLICATION NO.

EP 99 30 0740

This annex lists the patent family members relating to the patent documents cited in the above-mentioned European search report.  
The members are as contained in the European Patent Office EDP file on  
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

11-08-1999

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9520634 A	03-08-1995	AU 1599595 A	15-08-1995
		CA 2179907 A	03-08-1995
		CN 1139946 A	08-01-1997
		DE 69509365 D	02-06-1999
		EP 0741765 A	13-11-1996
		JP 9509196 T	16-09-1997
		US 5670557 A	23-09-1997
		US 5674561 A	07-10-1997
		US 5779632 A	14-07-1998
WO 9705171 A	13-02-1997	US 5853750 A	29-12-1998
		US 5670557 A	23-09-1997
		AU 6283796 A	26-02-1997
		CN 1191546 A	26-08-1998
		EP 0840753 A	13-05-1998
		US 5779632 A	14-07-1998
		US 5853750 A	29-12-1998
EP 0188381 A	23-07-1986	JP 1936150 C	26-05-1995
		JP 6068101 B	31-08-1994
		JP 61268767 A	28-11-1986
		US 4947847 A	14-08-1990
		US 4842768 A	27-06-1989
WO 8102097 A	06-08-1981	AU 543967 B	09-05-1985
		AU 6784081 A	17-08-1981
		BR 8009020 A	17-11-1981
		CA 1194647 A	01-10-1985
		DK 381081 A, B.	27-08-1981
		EP 0043850 A	20-01-1982
		IT 1142237 B	08-10-1986
		JP 1831926 C	29-03-1994
		JP 2174831 A	06-07-1990
		JP 3051413 B	06-08-1991
		US 4554924 A	26-11-1985
		US 4524087 A	18-06-1985
		US 4539996 A	10-09-1985
		ZA 8100460 A	24-02-1982
EP 0012402 A	25-06-1980	CA 1153427 A	06-09-1983
		CA 1237229 C	24-05-1988
		FR 2443843 A	11-07-1980
		JP 1436057 C	25-04-1988
		JP 55081635 A	19-06-1980
		JP 62044933 B	24-09-1987
		US 4391278 A	05-07-1983

EPO FORM P0459

For more details about this annex : see Official Journal of the European Patent Office, No. 12/82

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